

1 **Ocean acidification alters zooplankton communities and increases top-down pressure of a**
2 **cubozoan predator**

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23 **ABSTRACT**

24 The composition of local ecological communities is determined by the members of the regional
25 community that are able to survive the abiotic and biotic conditions of a local ecosystem. Anthropogenic
26 activities since the industrial revolution have increased atmospheric CO₂ concentrations, which have in
27 turn decreased ocean pH and altered carbonate ion concentrations: so called ocean acidification (OA).
28 Single-species experiments have shown how OA can dramatically affect zooplankton development,
29 physiology and skeletal mineralization status, potentially reducing their defensive function and altering
30 their predatory and anti-predatory behaviours. This means that increased OA may indirectly alter the
31 biotic conditions by modifying trophic interactions. We investigated how OA affects the impact of a
32 cubozoan predator on their zooplankton prey, predominantly Copepoda, Pleocyemata, Dendrobranchiata,
33 and Amphipoda. Experimental conditions were set at either current ($p\text{CO}_2$ 370 μatm) or end-of-the-
34 century OA ($p\text{CO}_2$ 1100 μatm) scenarios, crossed in an orthogonal experimental design with the
35 presence/absence of the cubozoan predator *Carybdea rastoni*. The combined effects of exposure to OA
36 and predation by *C. rastoni* caused greater shifts in community structure, and greater reductions in the
37 abundance of key taxa than would be predicted from combining the effect of each stressor in isolation.
38 Specifically, we show that in the combined presence of OA and a cubozoan predator, populations of the
39 most abundant member of the zooplankton community (calanoid copepods) were reduced 27 % more
40 than would be predicted based on the effects of these stressors in isolation, suggesting that OA increases
41 the susceptibility of plankton to predation. Our results indicate that the ecological consequences of OA
42 may be greater than predicted from single-species experiments, and highlight the need to understand
43 future marine global change from a community perspective.

44

45

46 INTRODUCTION

47 The composition of ecological communities is determined by the members of the regional species pool
48 that are able to pass through a series of ecological filters (Lawton 2000). Initially, to become part of a
49 community, species from the regional pool have to overcome the “dispersal filter”, i.e. they need to
50 physically enter the local community. Having arrived, the species then has to pass through the “abiotic
51 filter”, i.e. it has to be physiologically capable of tolerating the abiotic conditions present in the local
52 area. After clearing the dispersal and abiotic filters, a species must finally clear the “biotic filter”. In
53 order to pass through the biotic filter a species must be able to co-exist with the other members of the
54 regional species pool, species who themselves have already been able to pass through both the dispersal
55 and abiotic filters. Co-existing with these other community members means a species must not be
56 competitively excluded, or exploited to extinction by predators. It is impossible for a species to exert
57 competitive or consumptive pressure (i.e. become part of the biotic filter) if it cannot pass through the
58 dispersal or abiotic filters; thus, the filters concept is often depicted as a linear process (Fig. 1, Lawton
59 2000). Although the filters concept was first proposed to describe the presence or absence of species,
60 community composition can also change through changes in species’ relative abundances (Clarke, 1993;
61 Roemmich & McGowan, 1995; Forbes & Hammill, 2013). Changes in the strength of the different filters
62 may not necessarily lead to the exclusion of species, but community composition may still be altered
63 through reductions in their abundance.

64 Marine zooplankton communities are often dominated by Copepoda. Of the copepods, calanoids
65 are the most abundant and alone can constitute up to 80 % of total zooplankton biomass (Beaugrand *et*
66 *al.*, 2002; Lavaniegos & Ohman, 2007). The dominance of calanoid copepods, combined with the size of
67 the world’s oceans, suggests that they constitute the largest biomass of a single group of organisms on
68 earth and play a pivotal position in the food web (Gallienne & Robins, 2001; Stibor *et al.*, 2004; Turner,

69 2004). This high biomass makes marine copepods a crucial link between the ocean's primary producers
70 and higher trophic levels (Garzke *et al.*, 2016). This crucial role within pelagic marine food webs means
71 that any reduction in copepod abundance could decrease the survival of their consumers (Cross *et al.*,
72 2009), and have far-reaching consequences for marine communities (Frederiksen *et al.*, 2006).

73 The biological consequences of increased $p\text{CO}_2$ levels (ocean acidification - OA) in the marine
74 environment appear detrimental for many species (Fabry *et al.* 2008, Byrne *et al.* 2011, Kroeker *et al.*
75 2013, Przeslawski *et al.* 2015). Laboratory experiments and *in situ* observations at high $p\text{CO}_2$ sites (e.g.
76 CO_2 vents) have revealed that for calcifying organisms such as echinoderms, molluscs, corals, and
77 crustaceans, OA can broadly lead to reduced survival and developmental success (Hall-Spencer *et al.*,
78 2008; Sheppard Brennan *et al.*, 2010; Gaylord *et al.*, 2011; Kroeker *et al.*, 2011) and alter shell
79 mineralization (Byrne *et al.*, 2013; Wolfe *et al.*, 2013; Taylor *et al.*, 2015). However, some of the most
80 abundant and ecologically important species (e.g. copepods) in pelagic environments possess chitinous
81 exoskeletons that will not be as impacted by the dissolution status of sea water compared to calcifying
82 organisms (Kroeker *et al.*, 2013). Nevertheless OA has been associated with reduced adult and juvenile
83 survival (Lewis *et al.*, 2013; Cripps *et al.*, 2014), altered reproductive rates (Fitzer *et al.*, 2012; Thor &
84 Dupont, 2015), and reduced growth (Fitzer *et al.*, 2012) in copepods. These changes may be due to
85 changes in copepod metabolism in response to OA (Thor & Dupont, 2015), which then lead to the
86 reallocation of resources away from reproduction. These detrimental effects of OA on chitinous species
87 suggest that although these taxa may not suffer the same impacts on shell morphology, they are not
88 immune to the negative effects of OA.

89 The total effects of OA on individual taxa may also be magnified through indirect interspecific
90 interactions, as shown in experiments using pairs of grazers and herbivores (Poore *et al.*, 2013), or
91 predators and prey (Nagelkerken & Munday, 2016). Across a range of different biomes, changes in

92 pairwise consumer-resource interactions have been shown to scale up to the community level, leading to
93 changes in overall composition (Estes & Palmisano, 1974; Osman & Whitlatch, 2004; Hammill *et al.*,
94 2015a). Within marine ecosystems cubozoa (“box jellyfish”), are zooplanktivorous predators that
95 consume large volumes of zooplankton, including copepods (Buskey, 2003). Previous investigations into
96 the effects of OA on cubozoa have shown that they are capable of surviving and asexually reproducing at
97 pH levels as low as 7.6, although rates of asexual propagation through budding are reduced (Klein *et al.*,
98 2014). The ability of cubozoans to survive and reproduce (albeit at a reduced rate) under OA conditions
99 suggests an ability to tolerate potential future ocean conditions. What is not clear is whether OA reduces
100 the ability of cubozoans to capture prey, therefore reducing the effect of predation, or whether the
101 physiological effects of OA on their zooplankton prey makes escape harder, therefore increasing the
102 effect of predation. In terms of the ecological filters proposed by Lawton (Lawton, 2000), potential
103 changes in predation strength may lead to OA having indirect effects on the strength of the biotic filter,
104 potentially increasing the impact of OA on community structure (Fig. 1).

105 Here we investigate whether future OA conditions will alter plankton communities by changing
106 the strength of the abiotic filter, or by also increasing predation risk, thereby indirectly increasing the
107 strength of the biotic filter (Lawton, 2000). Using marine zooplankton communities and a natural
108 predator, a cubozoan jellyfish, we conducted a microcosm experiment in which the presence of predators
109 and OA was crossed in a 2 x 2 factorial design: $p\text{CO}_2$ 370 or 1100 μatm crossed with predator
110 presence/absence. We predicted that in isolation, OA conditions or the presence of cubozoan predators
111 would alter the composition of zooplankton communities by reducing the abundances of multiple taxa,
112 including dominant copepods. However, we also predicted that when OA and predators were present
113 simultaneously, their combined impacts on community structure, driven by changes in copepod

114 abundances would be greater than would be predicted from combining their isolated effects, as OA is
115 likely to increase susceptibility to predation.

116

117 **MATERIALS AND METHODS**

118 *Organism collections*

119 Zooplankton were collected from Chowder Bay in Sydney Harbour, Australia (-33° 50' 59.6394", 151°
120 15' 0.36") using a 250 µm plankton net. Many zooplankton taxa swarm in the presence of light, a
121 behavioral tactic that has been exploited by cubozoans (Buskey, 2003). Plankton were therefore collected
122 at night by illuminating the water surface using 200 lumen lamps (Icon© Headlamp, Black Diamond
123 Equipment LTD, Salt Lake City, UT, USA), then towing the plankton net 20 times through the
124 illuminated water. After each tow the contents of the net were emptied into a 20 L white plastic
125 bucket. The net was emptied carefully after each tow to minimize damage to organisms. While
126 illumination can alter the relative abundances of zooplankton, a previous investigation in temperate
127 waters showed that the use of illumination led to the exclusion of only 1 out of 43 taxa, as opposed to
128 excluding 9 out of 43 taxa in the absence of illumination (McConnell *et al.*, 2010). We therefore opted to
129 use illumination as it facilitated rapid collection of large numbers of the most abundant taxa, due to the
130 positive phototaxis characteristic of many zooplankton (Buskey, 2003; Porter *et al.*, 2008; Martynova &
131 Gordeeva, 2010). This method also substantially reduced the handling time needed to obtain sufficient
132 populations for the experiment. Following collection, fresh sea water was added to the bucket to make
133 the volume up to 15 L. The contents of the bucket were then thoroughly mixed through gentle stirring for
134 30 sec, and divided into 75 subsamples each of 200 mL. Twelve plankton kreisels were then inoculated
135 with four of the 200 mL subsamples, selected at random. We did not identify the organisms introduced
136 into each kreisel prior to the start of the experiment to avoid stress associated with microscopic

137 identification. To assess the initial composition of the plankton, we collected a second zooplankton
138 sample that was handled in an identical manner (eg. collection, 75 x 200 mL subsamples), where the
139 subsamples were immediately fixed in 75 % ethanol for later identification. These preserved subsamples
140 were identified to ascertain initial composition and to quantify the likelihood that significant differences
141 existed among the experimental replicates at the start of the experiment. After the zooplankton
142 communities within these subsamples were quantified, we performed numerical simulations in which
143 every possible combination of subsamples were randomly assigned to the different experimental kreisels,
144 and looked for significant differences among treatments. This simulation exercise revealed that there was
145 a < 5 % chance that significant differences existed among the treatments at the start of the experiment
146 (*Supplementary Materials*).

147 The cubozoan *Carybdea rastoni* naturally occurs in and around Sydney Harbour (The
148 Australian Museum, 2016). *Carybdea rastoni* have a broad diet consisting of zooplankton and small fish
149 (Larson, 1976). Due to its ease of identification and capture, we used this species as the gelatinous
150 predator in the experiment. Because of their delicate body structure, *C. rastoni* were individually
151 collected from the water using a 2 L bucket rather than a plankton net. For the treatments containing
152 predators, a single individual *C. rastoni* was placed in each experimental kreisel at the start of the
153 experiment. All *C. rastoni* used in the experiment measured between 20 and 25 mm across the widest
154 part of the bell, and we found no differences in size among treatments ($t_{(4)} = 0.35$, $P = 0.74$). We were
155 able to capture sufficient *C. rastoni* for the experiment in less than 1 hour, suggesting they were relatively
156 abundant in Chowder Bay at the time of collection.

157

158 *Experimental vessels*

159 Custom-built plankton kreisels (Greve, 1968) were used as the experimental vessels (n = 12). Plankton
160 kreisels allow pelagic zooplankton to be cultured in flow-through systems, while preventing individuals

161 adhering to solid surfaces. Each kreisel consisted of a tank measuring 260 mm x 400 mm x 73 mm, with
162 the circular component being a 73 mm-wide section of PVC tubing with a diameter of 210 mm (Fig. S1).
163 Placing the circular component within the tank generates a circular current, preventing adhesion of
164 zooplankton. A “slip” was inserted immediately adjacent to the inflow in order to create a more laminar
165 flow. An outflow tube was installed in the circular component and covered with 100 µm mesh to prevent
166 any organisms escaping. We used three kreisels in each of the four experimental treatments, and the
167 experiment was ran for 10 d. Throughout the experiment, 150 µL of Instant Algae (Shellfish Diet 1800[®],
168 Reed Mariculture, Campbell CA, USA) was added to each kreisel daily. As the water was only filtered to
169 20 µm, the flow through system also supplied phytoplankton as an additional source of food (Byrne,
170 *pers. obs.*).

171

172 *Experimental conditions*

173 The kreisels were supplied with filtered sea water (FSW; 20 µm filtered) from four 60 L header tanks.
174 Manipulating $p\text{CO}_2$ within these four header tanks allowed them to supply different OA treatments to the
175 experimental kreisels. Two header tanks were assigned as containing ambient sea water ($p\text{CO}_2$ 370
176 µatm), and two contained elevated $p\text{CO}_2$ sea water ($p\text{CO}_2$ 1100 µatm, see table 1). Each header tank
177 consisted of a 60 L plastic tub with a tight fitting lid. Filtered sea water entered each of the header tanks
178 at a rate of 1 L min⁻¹. The OA treatment was achieved by bubbling CO₂-enriched air vigorously through
179 sea water using 20 cm ceramic diffusers. In the two header tanks supplying current conditions, ambient
180 air was bubbled through the diffusers. To achieve target $p\text{CO}_2$ for the OA treatments, ambient air was
181 scrubbed of CO₂ and then mixed with industrial-grade CO₂ (BOC, Sydney, Australia) through a VSO[®]
182 thermally compensated low flow controller valve (Parker Hannifin, USA). A software-controlled,
183 proportional–integral–derivative (PID) device ensured $p\text{CO}_2$ concentration in the gas mix at the level

184 predicted for the year 2050 (CSIRO and Bureau of Meteorology 2016). Header tank temperatures were
185 maintained using software-controlled solenoid valves that mixed warm and cold water to achieve a
186 constant temperature of 21 °C, representing ambient temperature at the time of collection of the plankton.
187 Each header tank then supplied three of the experimental kreisels, *via* gravity, at a rate of 300 mL min⁻¹.
188 Excess water within the header tank drained out via an overflow.

189 Treatment stability was monitored every second day using a handheld multimeter (Multi 3430 Set G,
190 Wissenschaftlich-Technische Werkstätten GmbH, Weilheim, Oberbayern, Germany) equipped with a
191 temperature/pH sensor (SenTix 940, Wissenschaftlich-Technische Werkstätten GmbH). On each
192 monitoring occasion, pH measurements were taken from each of the three experimental kreisels. The pH
193 electrode was calibrated prior to use with high precision NIST buffers pH 4, 7 and 10 (ProSciTech,
194 Townsville, QLD, Australia). Total alkalinity (TA) was determined from water samples collected every
195 second day through potentiometric titration (907 Titrando, Metrohm) using certified reference standards
196 (Dickson et al., 2007). Full physico-chemical parameters including bicarbonate and carbonate ion
197 concentration ($[\text{HCO}_3^-]$ and $[\text{CO}_3^{2-}]$) and calcite and aragonite saturation states (Ω_{cal} and Ω_{ara}) were
198 subsequently calculated using CO₂SYS (Pierrot *et al.*, 2006), using CO₂ dissociation constants K_1 and
199 K_2 from Mehrbach et al. (1973) refit by Dickson and Millero (1987), and K_{SO_4} from Dickson et al.
200 (2007). Values are provided in Table 1. Within each of the OA treatments, predators were randomly
201 allocated to the three kreisels being fed by each of the header tanks. As each OA treatment was fed by
202 two header tanks, each of which supplied water to three kreisels, one header tank would supply two
203 predator replicates, and the other would supply one. A coin toss was used to establish which of the header
204 tanks in each OA treatment would supply two predator replicates. Following this initial allocation, the
205 individual kreisels that would house communities containing predators were identified using a random
206 number generator. This process was undertaken to produce a random allocation of predators across

207 header tanks, while also ensuring one header tank in each OA treatment did not also supply all predator
208 treatments.

209 *Data collection, visualization, and Statistical Analyses*

210 At the termination of the experiment, all zooplankton were collected from each of the experimental
211 kreisels and preserved in 75 % ethanol. Zooplankton abundances were calculated by identifying each
212 individual using a Leica EZ4 stereo dissecting microscope. Differences in zooplankton community
213 composition between treatments were visualized using multidimensional scaling (MDS) plots, a method
214 that implements ordination methods to illustrate differences in multidimensional data. Abundance of each
215 zooplankton taxon was used in the MDS plots, and visualized in 2-dimensions. Within this 2-
216 dimensional space, communities that have compositions similar to each other are clustered together,
217 whereas communities with different compositions are spaced further apart. Formal analysis of differences
218 in the zooplankton community composition data required a simultaneous analysis of multiple response
219 variables (abundances of each zooplankton taxa) using two explanatory variables (OA and predator
220 presence). We therefore quantified differences in overall community composition using PERMANOVA
221 within the vegan package, using the R statistical programming language (R Foundation for Statistical
222 Computing, 2015). PERMANOVA analyses incorporate distance matrices, and are considered a robust
223 metric to analyze differences in community composition (Forbes & Hammill, 2013; Hammill *et al.*,
224 2015a). As our data had two independent descriptive variables (OA and presence/absence of a cubozoan
225 predator), we performed a 2-way PERMANOVA including an interaction term. The contributions of
226 different taxa to community level differences were calculated using SIMPER (Warton *et al.*, 2012).

227 While PERMANOVA indicates differences in multivariate data, it does not describe how
228 communities differ, or demonstrate if a significant interaction represents an antagonistic or synergistic
229 effect. To better understand how community composition is altered due to OA and predator presence, we

230 performed 2-way ANOVAs on each zooplankton taxon to analyze differences in abundance. Within each
231 of the ANOVA tests, we included an OA x predator interaction term. To understand whether a significant
232 interaction term indicated an antagonist or synergistic interaction between OA and predators, we
233 generated multiplicative risk models (Soluk & Collins, 1988; Atwood *et al.*, 2014), then compared the
234 result of these null models to our observed data. The general form of the multiplicative risk null model
235 (MRNM) is described by the following equation:

236

237 (Eq. 1)
$$MRNM = (E_{OA} + E_{predators}) - (E_{OA} * E_{predators})$$

253

238 where *MRNM* is the predicted effect of the combined stressors (i.e. the “null model” or the abundance we
239 would expect if predators and OA acted independently of each other), as a proportional change in the
240 metric of interest (e.g. the abundance of each zooplankton). E_{OA} and $E_{predators}$ represent the respective
241 observed proportional changes due to OA and the presence of a cubozoan predator. These multiplicative
242 null models are useful as they never predict reductions in abundances greater than 100%. Should our
243 observed reductions in zooplankton abundance due to both OA and the presence of predators be greater
244 than predicted by the null model, this would indicate these two factors act synergistically. If reductions in
245 abundance are less than predicted by a null model, this indicates an antagonistic interaction. We also
246 performed numerical simulations for the taxa where a significant OA x predator interaction was found to
247 compare predator consumption rates under ambient and elevated pCO_2 conditions (*Supplementary*
248 *Materials*).

249 Within the PERMANOVA and ANOVA tests, “header tank ID” was included as a blocking term.
250 In all analyses, we found no significant effect of header tank ID, and no significant interactions with
251 either predator presence/absence or OA condition (all $P > 0.05$). We therefore removed all terms
252 involving header tank ID and re-ran the analyses.

254

255 **Results**

256 *Plankton community*

257 At the end of the experimental trials, all members of the zooplankton community belonged to the
258 subphylum Crustacea. This dominance of Crustacea is consistent with the analysis of the initial
259 community composition, and all taxa appeared to survive well in the kreisels during the experiment
260 (*Supplementary Materials*). Adult copepods were the most abundant members of the community,
261 accounting for 91.8 % of the total number of individuals. Of these copepods, the vast majority (99.6 %)
262 were members of the order Calanoida, with the remainder being members of the order Cyclopoida. After
263 adult copepods, the second most abundant members of the community were copepod nauplii (3.1 %). The
264 remainder of the community consisted of members of the orders Amphipoda (2.5 %), Pleocymata (1.7
265 %), Dendrobranchiata (0.6 %) and Mysida (0.4 %).

266

267 *Community-level effects of experimental stressors*

268 We found that community-level changes in the presence of both drivers were different from what would
269 be expected given the results of these stressors in isolation, as indicated by the presence of a significant
270 OA x predator interaction ($F_{(1,8)} = 28.99$, $P < 0.001$, PERMANOVA, Fig. 2). In isolation, OA was
271 associated with significant changes in community composition ($F_{(1,8)} = 59.81$, $P < 0.001$, Fig. 2). The
272 addition of a cubozoan predator in isolation also led to significant changes in the zooplankton community
273 ($F_{(1,8)} = 27.34$, $P < 0.001$, PERMANOVA, Fig. 2). Within the zooplankton community, calanoid
274 copepods were the greatest contributor to differences between treatments, accounting for 88.52 ± 2.30 %
275 of the differences between treatments (SIMPER).

276

277 *Effects of experimental drivers on individual zooplankton taxa*

278 Reductions in calanoid abundances due to a combination of cubozoan predation and OA were $27.00 \pm$
279 1.50% greater than would be expected based on the results of these drivers in isolation (OA x predator
280 interaction, $F_{(1,8)} = 45.61$, $P < 0.001$, ANOVA, Fig. 3a). Predation rates appeared to increase under OA
281 conditions, with cubozoan predators consuming 36.7% (95% CI: $23.02\% - 50.08\%$) of available
282 calanoids at ambient $p\text{CO}_2$, and 82.7% (95% CI: $62.50\% - 97.98\%$) under elevated $p\text{CO}_2$
283 (*Supplementary Materials*). Abundances of pleocyematids were also reduced by $61.34 \pm 7.80\%$ more
284 through a combination of cubozoan predators and OA than would be predicted by the effect of these
285 stressors in isolation, demonstrated by a significant interaction term and comparison against the
286 multiplicative null model ($F_{(1,8)} = 6.25$, $P = 0.033$, Fig. 3b). Cubozoan predators consumed 0% (95% CI:
287 $-28.60\% - 28.60\%$) of pleocyematids at ambient $p\text{CO}_2$, but the rate of consumption increased to 71.5%
288 (95% CI: $42.09\% - 100.1\%$) under elevated $p\text{CO}_2$ (*Supplementary Materials*). These results indicate that
289 predation rates on calanoids and pleocyematids increased under OA conditions.

290 In addition to the significant OA x predator interactions, abundances of certain zooplankton taxa
291 were also reduced by each of our experimental drivers when applied in isolation. Abundances of
292 calanoids were $59.4\% \pm 8.6\%$ lower under OA conditions compared to controls ($F_{(1,8)} = 251.65$, $P <$
293 0.001 , ANOVA Fig. 3a). In addition, copepod nauplii were completely absent under OA conditions ($F_{(1,8)}$
294 $= 110.09$, $P < 0.001$, Fig. 3c). Under current ambient conditions, the presence of a cubozoan predator led
295 to a $36.6\% \pm 9.5\%$ reduction in the abundance of calanoids ($F_{(1,8)} = 127.46$, $P < 0.001$, ANOVA Fig.
296 3a). Dendrobranchids were also entirely absent in the presence of cubozoan predators, in both ambient
297 and OA conditions ($F_{(1,9)} = 12.00$, $P = 0.007$, Fig. 3d).

298 For all other zooplankton orders and sub-orders (Cyclopoida, Amphipoda, and Mysida), we found
299 no significant changes in abundance following exposure to OA conditions, the presence of cubozoan
300 predators, or an interaction between the two (all $P > 0.05$, Fig. 3e-g). All cubozoan predators survived,
301 and were observed to be freely moving around the kreisels at the termination of the experiment.

302

303 **Discussion**

304 Our data revealed that ocean acidification (OA) can lead to substantial changes in the composition of
305 zooplankton communities through direct and indirect mechanisms. Crucially, when compared to
306 multiplicative null models, the changes in abundance we observed for multiple taxa under a combination
307 of OA and predators were greater than would be expected based on the effects of these stressors in
308 isolation. This synergistic effect indicates that OA will not only directly affect community composition
309 by changing the strength of the abiotic filter, but also by altering the strength of the biotic filter through
310 increasing predation risk (Lawton, 2000).

311 In isolation, OA and the presence of cubozoan predators were associated with significant changes
312 in community composition, driven by reductions in calanoid copepods and copepod nauplii. These
313 changes in copepod abundance indicate that OA can alter the strength of the abiotic filter to such a degree
314 that it can detrimentally affect the most abundant member of the zooplankton community. As predicted,
315 under current conditions, exposure to cubozoan predators also altered the composition of the zooplankton
316 community, through reductions in abundances of Calanoida and Dendrobranchiata. These predator-
317 mediated changes in overall community composition indicate that cubozoan predators represent a current
318 and important component of the biotic filter.

319 Although the exoskeletons of chitinous organisms are less affected by OA than calcified taxa
320 (Cohen *et al.*, 2009; Byrne *et al.*, 2011; Chan *et al.*, 2011; Gaylord *et al.*, 2011), previous investigations

321 show that copepods can be affected through changes in metabolic rates (Thor & Dupont, 2015), which
322 can be compensated for by increased grazing rates (Li & Gao, 2012). These metabolic changes have been
323 associated with reallocation of energetic resources, and may lead to a reduction in the amount of energy
324 available for escape. Calanoid copepods have been shown to escape gelatinous predators by initiating
325 multiple escape jumps, which can lead to less than 1% of predator encounters resulting in capture
326 (Suchman, 2000). Because of this reliance on an energetically expensive escape strategy, any abiotic
327 change that reduces energy available for escape could increase predation on copepods. We specifically
328 provided an over-abundance of food to increase the chances that individuals had sufficient resources for
329 growth and reproduction. However, we cannot rule out that under OA conditions, changes to resource
330 requirements (Li & Gao, 2012), and metabolism (Thor & Dupont, 2015) limited the ability of
331 zooplankton to survive in the presence of a predator. Although untested here, this may represent a series
332 of additional physiological mechanisms by which OA affects copepod susceptibility to predation and
333 represents an important area for future investigations.

334 To our knowledge, no studies have been conducted on the foraging mode of *C. rastoni*. However,
335 similar species of cubozoan jellyfish are considered to be largely “semi-passive hunters” (although see
336 Courtney *et al.*, (2015)), using low resolution vision to seek out habitats with high prey densities and then
337 passively collecting prey on their extended tentacles as they swim (Buskey, 2003; Garm *et al.*, 2011).
338 This type of foraging mode most resembles filter feeding, which is characterized as having a Type I
339 functional response (Titelman & Hansson, 2006; Møller & Riisgård, 2007). Within a Type I functional
340 response, predators should consume a constant proportion of their prey regardless of prey density.
341 However, in the current study, cubozoan predators consumed a greater proportion of the available
342 copepods under OA conditions compared with ambient (*Supplementary Materials*), suggesting OA
343 increased the proportion of food available for capture. Conversely, OA may increase the food

344 requirements of cubozoans, causing them to dedicate more effort to prey collection. This increase in prey
345 collection effort could mean that copepods and other taxa need not become more susceptible to predation
346 under OA conditions. The increased susceptibility of prey to predation and increased food requirements
347 of predators may not be mutually exclusive, meaning that the observed reduction in copepods may be a
348 byproduct of their additive effect. We therefore cannot conclusively determine whether the increased
349 predation rates observed under OA conditions were the result of increased prey susceptibility, or
350 increased predation effort. Future analyses into how OA alters the metabolism and behavior of both
351 predators and prey may aid in teasing apart the individual-level or physiological-level mechanisms at the
352 base of our results, and provide insight into whether it is OA-mediated changes in predators, prey, or both
353 that are driving the patterns we observe here. Nonetheless, our results do demonstrate that OA increases
354 the top-down pressure exerted on zooplankton communities by individual cubozoan predators,
355 highlighting the importance of considering predator-dependent mortalities when considering the full
356 impacts of OA.

357 All *C. rastoni* survived until the experiment was terminated after 10 days, and were observed
358 freely moving around the kreisels. This 100% survival rate implies that *C. rastoni* are relatively tolerant
359 of OA conditions, at least in terms of survival over a relatively short time period. Physiological
360 investigations into the effects of OA on cubozoans have indicated that they are able to cope with near
361 future OA conditions, showing no significant changes in survival, and maintaining asexual reproduction
362 (although at a lower rate) (Klein *et al.*, 2014). The reduced reproduction rates observed under OA for
363 cubozoans in previous experiments (Klein *et al.*, 2014) may mean that although each individual cubozoan
364 exerts more top-down pressure, there are fewer cubozoans in the ecosystem. The total net effect of the
365 cubozoan population on their prey will therefore depend on whether the decrease in reproduction is
366 greater than, equal to, or less than the increase in predation rates. Short-term predator-prey experiments

367 such as ours provide insights into the factors that alter interaction strength, such as the relationships
368 between biodiversity and strength of trophic links (Kratina *et al.*, 2007). However, complementary
369 longer-term (> 2 predator generations) experiments are necessary to understand if these short-term
370 experiments translate to changes in overall community dynamics, rather than composition at a single time
371 point (compare Kratina *et al.* 2007 to Hammill *et al.* 2015b). As the experiment we present here was
372 conducted over a relatively short time frame (< 1 predator generation), we do not know if these changes
373 in the strength of trophic interactions translate into long-term alterations in the relative abundances of
374 predators and prey.

375 At the population level, several studies have reported OA affecting life history parameters that
376 influence population growth rates (Albright *et al.*, 2010; Fitzer *et al.*, 2012). Given the rapid generation
377 time of copepods (Gillooly, 2000), nauplii introduced at the start of our experiment could have reached
378 the adult stages by the end of the experiment. However, it is unlikely that our reduced copepod nauplii
379 abundances were caused by an increase in the number of individuals transitioning to the adult stages, as
380 OA was also associated with a reduction in the number of adult copepods. Furthermore, predation alone
381 did not decrease copepod nauplii. Our observation of reduced nauplii abundances under OA conditions is
382 therefore best explained by previously reported factors such as reduced reproduction (Fitzer *et al.*, 2012;
383 Thor & Dupont, 2015), reduced hatching success (Mayor *et al.*, 2007), or increased nauplii mortality
384 (Pedersen *et al.*, 2014). Given the substantial reduction in nauplii and adult copepods we observed, OA
385 could have significant consequences for the population trajectory of copepods, and their roles within
386 ecosystems. We observed no nauplii under elevated $p\text{CO}_2$, suggesting that copepod reproduction was
387 dramatically reduced, potentially causing their populations to crash. Copepods are a dominant member of
388 zooplankton communities (Beaugrand *et al.*, 2002; Lavaniegos & Ohman, 2007) and exert significant
389 grazing pressure on primary producers (Sommer & Sommer, 2006). A reduction in top-down control

390 associated with an OA-mediated drop in copepod populations, coupled with the ability of phytoplankton
391 to capitalize on the increased $p\text{CO}_2$ associated with OA (Schippers *et al.*, 2004), may lead to an increase
392 in magnitude of phytoplankton blooms under future ocean conditions.

393 The challenges that OA poses for the production and maintenance of heavily calcified skeletons
394 means that much previous research has focused on organisms possessing these structures (Gaylord *et al.*,
395 2011; Coleman *et al.*, 2014). While the chitinous skeletons of copepods and other planktonic crustaceans
396 may make this group less susceptible physiologically to OA (Poore *et al.*, 2013), the large decrease in
397 copepod nauplii production we observed could lead to substantial reductions in copepod populations.
398 Copepods have been proposed to account for the highest biomass of a single group of organisms on earth
399 (Gallienne & Robins, 2001; Turner, 2004), and also form the important link between pelagic primary
400 producers and economically important fisheries (Frederiksen *et al.*, 2006). Reductions in copepod
401 numbers have also been associated with declines in members of higher trophic levels such as salmon
402 (Cross *et al.*, 2009), highlighting that OA-mediated changes low in the food chain could influence many
403 species in marine food webs. Understanding how OA affects lower trophic levels in a community-wide
404 context is therefore critical to elucidating the total effects of changing oceanic conditions.

405 While our experiment indicates that zooplankton communities are altered through a combination
406 of OA and predators, it was designed and conducted as a proof of concept. As our study was conducted in
407 a single geographic location, and at only one point in time, we are cautious about generalizing our results
408 to a wider geographic area. We can, however, speculate about how the geographic location affected our
409 experiment, and how we may expect the results to differ in other areas. Previous studies indicate that
410 species experiencing fluctuating conditions have been shown to be more tolerant to changes in abiotic
411 conditions than species that experience greater stability (Maas *et al.*, 2012; Lewis *et al.*, 2013). The
412 temperate nature of Chowder Bay is associated with annual fluctuations in mean temperatures between

413 17 and 25 °C (<http://www.seatemperature.org/>). While data on pH fluctuations in Chowder Bay are not
414 available, the near shore, temperate nature of the region would suggest that pH levels are variable
415 (Hofmann *et al.*, 2011). These fluctuations in pH may provide a buffering effect on zooplankton
416 communities, we therefore need to understand the interplay between magnitudes of OA and the
417 susceptibility of different communities to determine the full community-level consequences of OA on
418 marine communities.

419 We have demonstrated that OA may directly increase the strength of the abiotic filter, and
420 indirectly increase the strength of the biotic filter in a marine plankton community. As a consequence of
421 the indirect increase in the strength of the biotic filter, OA may not need to cause mortality in isolation to
422 substantially change the composition of pelagic communities. Our results highlight the need to consider
423 direct and indirect effects if we are to understand the total impacts of OA on marine communities in the
424 coming decades (Kroeker *et al.*, 2013; Poore *et al.*, 2013; Gaylord *et al.*, 2015).

425

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433

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588 *erythrogramma*). *Global Change Biology*, **19**, 2698–2707.

589

| Header Tank | Ambient treatment | | | | | | OA treatment | | | | | |
|---|-------------------|---------|---------|---------|---------|---------|--------------|---------|---------|---------|---------|---------|
| | 1 | | | 2 | | | 3 | | | 4 | | |
| Predator present | Yes | No | No | Yes | No | Yes | Yes | Yes | No | No | No | Yes |
| Kreisel number | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
| Temperature (°C) | 21.11 | 21.09 | 21.08 | 21.11 | 21.06 | 21.14 | 21.08 | 21.1 | 21.09 | 21.1 | 21.11 | 21.12 |
| (±S.E) | 0.1 | 0.07 | 0.09 | 0.05 | 0.05 | 0.1 | 0.07 | 0.08 | 0.08 | 0.06 | 0.06 | 0.07 |
| Salinity | 34.62 | 34.68 | 34.65 | 34.66 | 34.64 | 34.68 | 34.65 | 34.67 | 34.65 | 34.64 | 34.65 | 34.63 |
| (±S.E) | 0.04 | 0.06 | 0.05 | 0.03 | 0.05 | 0.05 | 0.05 | 0.06 | 0.06 | 0.07 | 0.06 | 0.07 |
| pH _{NIST} | 8.20 | 8.23 | 8.22 | 8.19 | 8.20 | 8.19 | 7.79 | 7.79 | 7.8 | 7.77 | 7.77 | 7.77 |
| (±S.E) | 0.02 | 0.04 | 0.02 | 0.03 | 0.02 | 0.03 | 0.02 | 0.03 | 0.03 | 0.05 | 0.05 | 0.05 |
| TA (μEq kg ⁻¹) ^a | 2294.96 | 2295.64 | 2293.39 | 2298.66 | 2295.11 | 2297.55 | 2290.43 | 2298.74 | 2293.47 | 2290.37 | 2296.11 | 2294.16 |
| (±S.E) | 10.35 | 10.18 | 5.19 | 7.53 | 4.57 | 6.78 | 9.5 | 5.43 | 6.7 | 9.65 | 14.55 | 7.97 |
| pCO ₂ (μatm) ^a | 367.81 | 356.1 | 353.6 | 379.16 | 367.76 | 364.2 | 1096.82 | 1090.78 | 1079.88 | 1161.53 | 1157.7 | 1157.56 |
| (±S.E) | 22.49 | 22.5 | 16.28 | 27.02 | 23.38 | 37.95 | 56.38 | 87.37 | 88.44 | 90.2 | 83.84 | 84.17 |
| DIC (μmol kg ⁻¹) ^a | 2013.82 | 2001.72 | 2004.79 | 2012.6 | 2014.08 | 2013.4 | 2195.45 | 2192.62 | 2195.62 | 2204.28 | 2207.86 | 2205.92 |
| (±S.E) | 17.01 | 16.37 | 9.89 | 17.03 | 11.98 | 18.84 | 12.92 | 15.45 | 15.21 | 13.85 | 16.12 | 12.54 |
| HCO ₃ ⁻ (μmol kg ⁻¹) ^a | 1801.97 | 1782.39 | 1788.72 | 1803.57 | 1802.33 | 1805.46 | 2072.23 | 2069.02 | 2071.3 | 2083.24 | 2082.01 | 2084.14 |
| (±S.E) | 22.81 | 23.24 | 15.37 | 24.45 | 19.41 | 29.37 | 14.01 | 18.38 | 17.97 | 15.4 | 16.9 | 14.41 |
| CO ₃ ²⁻ (μmol kg ⁻¹) ^a | 200.29 | 208.46 | 204.97 | 197.13 | 200.19 | 195.89 | 88.76 | 89.36 | 90.39 | 84.27 | 85.51 | 85.47 |
| (±S.E) | 7.74 | 8.98 | 6.59 | 8.92 | 8.51 | 12.21 | 3.73 | 5.82 | 5.92 | 5.22 | 5.13 | 5.26 |
| Ω _{cal} ^a | 4.81 | 5 | 4.92 | 4.83 | 4.81 | 4.8 | 2.13 | 2.15 | 2.17 | 2.02 | 2.05 | 2.05 |
| (±S.E) | 0.19 | 0.22 | 0.16 | 0.21 | 0.21 | 0.29 | 0.09 | 0.14 | 0.14 | 0.13 | 0.12 | 0.13 |
| Ω _{ara} ^a | 3.13 | 3.16 | 3.21 | 3.08 | 3.13 | 3.06 | 1.39 | 1.4 | 1.41 | 1.35 | 1.34 | 1.34 |
| (±S.E) | 0.12 | 0.14 | 0.1 | 0.14 | 0.13 | 0.19 | 0.06 | 0.09 | 0.09 | 0.08 | 0.08 | 0.08 |

592

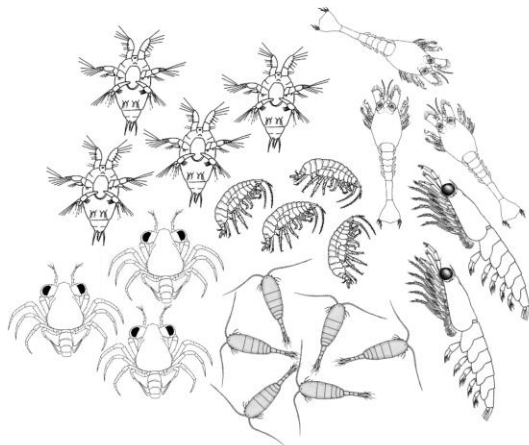
593 **Table 1.** Physico-chemical parameters of kreisels used to assign ocean acidification (OA) treatments (mean ± SE): temperature (°C),
594 pH (NIST scale), salinity, total alkalinity (TA μEq kg⁻¹), dissolved inorganic carbon (DIC μmol kg⁻¹), carbon dioxide partial pressure
595 (pCO₂ μatm), bicarbonate concentration (HCO₃⁻ μmol kg⁻¹), carbonate concentration (CO₃²⁻ μmol kg⁻¹), calcite (Ω_{cal}) and aragonite
596 saturation (Ω_{ara}). Header tanks 1 and 2 supplied sea water at current oceanic conditions, header tanks 3 and 4 represent OA conditions.
597 All ionic concentrations were calculated in CO2SYS (Pierrot *et al.*, 2006), using CO₂ disassociation constants K₁ and K₂ from
598 Mehrbach *et al.* (1973) refit by Dickson and Millero (1987), and KSO₄ from Dickson *et al.* (2007)

599 **Figure Legends**

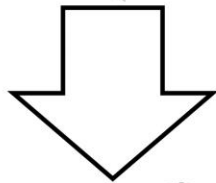
600 **Fig. 1.** Illustration of the filters that determine the composition of a local community. The
601 dispersal filter is not included as it did not form part of the current experiment. The abundances
602 of species present in the regional pool represent the potential community. The members of the
603 regional pool that are able to cope with ocean acidification and a cubzoan predator in
604 combination determine the final community composition. Changes in OA may alter final
605 community composition by directly affecting the strength of the abiotic filter, or by increasing
606 susceptibility to predation, thereby increasing the strength of the biotic filter. After Lawton
607 (2000).

608 **Fig. 2.** Multidimensional scaling plot illustrating how OA and the presence/absence of predators
609 alters community composition. Each point represents the community contained within a single
610 plankton kreisel at the end of the experiment. Distances between points are proportional to
611 similarities in community composition, meaning nearby points represent similar communities.

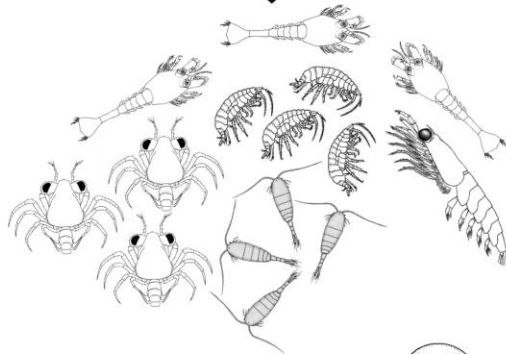
612 **Fig. 3.** Consequences of (ocean acidification) OA and predator presence/absence on the Log₁₀
613 abundances (± 1 standard error) of zooplankton within the community. **a)** Calanoida copepods,
614 **b)** Pleocyemata, **c)** Nauplii of all copepods, **d)** Dendrobranchiata, **e)** Amphipoda, **f)** Mysida, **g)**
615 Cyclopoida copepods. Different letters above bars indicate treatments that are significantly
616 different from each other. In each panel, the black “null model” bar represents the predicted
617 additive effects of OA and scyphozoan predators if these two factors acted independently, and
618 asterisks indicate synergistic interaction



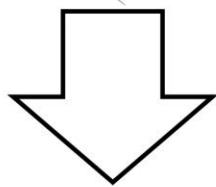
Potential community composition, determined by regional pool



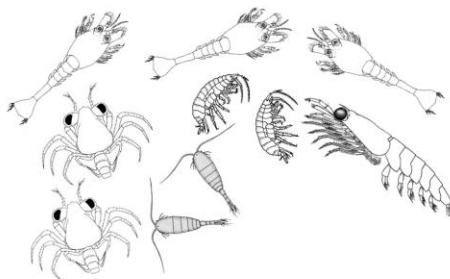
Abiotic filter (altered pH)



Community able to overcome abiotic conditions



Biotic filter (cubozoan predator)



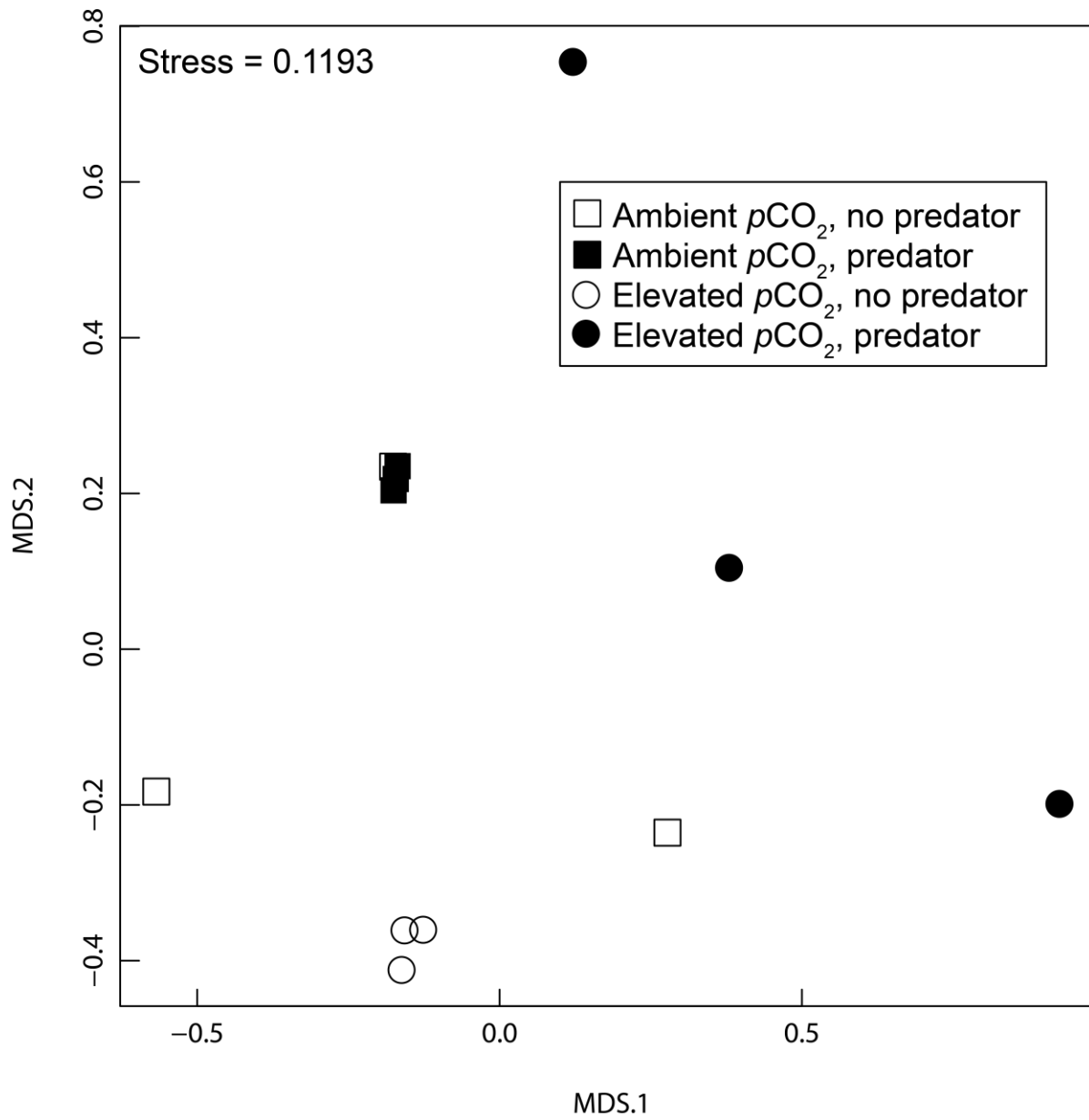
Community able to overcome abiotic and biotic conditions

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622

623 Figure 2



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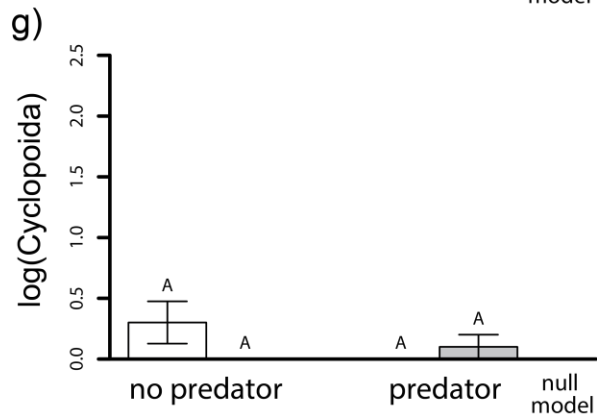
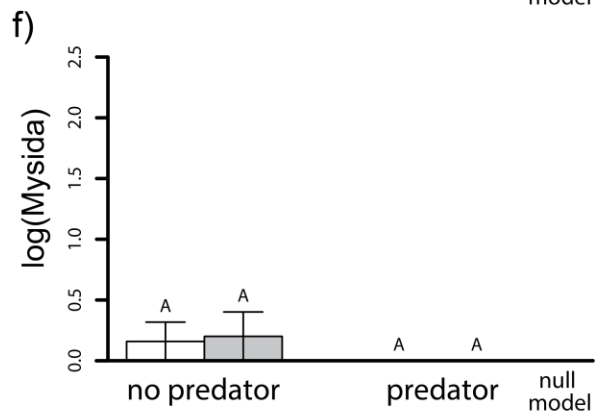
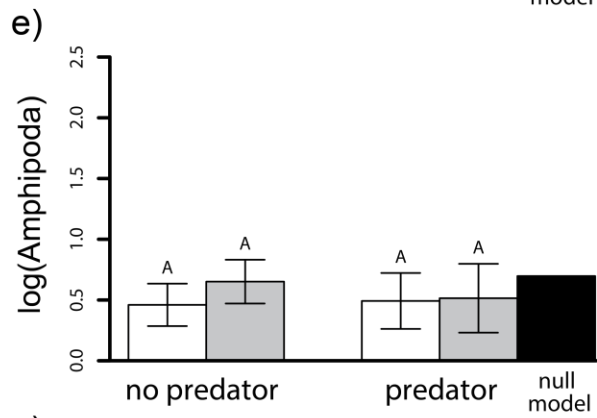
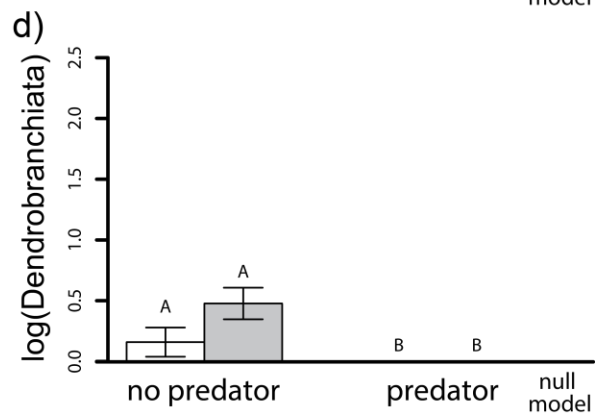
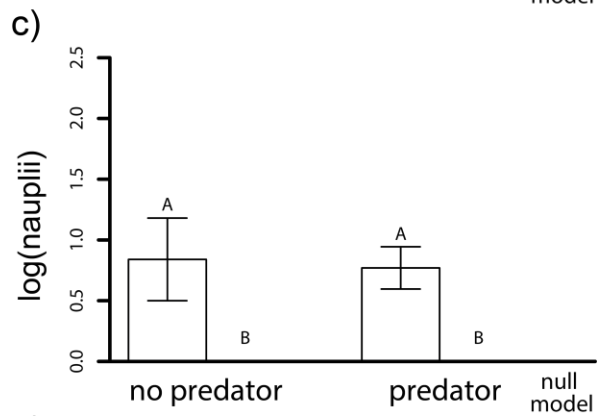
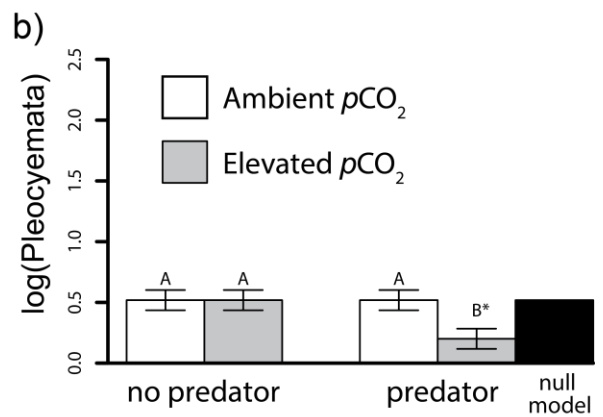
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632 Figure 3



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